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# Skeletal ontogeny of the caudal complex in Caspian kutum, *Rutilus kutum* (Kamensky, 1901) (Teleostei: Cyprinidae) during early development

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# ABSTRACT

Study of skeletal ontogeny during early development is crucial in a successful larviculture. In this study, ontogeny of the caudal complex skeleton in Caspian kutum, *Rutilus kutum*, was studied from hatching up to 50-day post hatching (dph). A total of 520 larvae and fry were randomly sampled, cleared and stained for osteological examinations. The results showed that development of caudal skeleton is started with appearing hypurals 1 and 2 on 4 dph and inflexion occurred on 7 dph with formation of eight cartilaginous plates, including two haemal spines, parhypural and five hypurals, below the notochord. The results also revealed that during post flexion from 13 to 50 dph other structures e.g. rudimentary neural arch, second ural and anterior half of the neural arch of PU3 were appeared. The inflexion point of tail growth pattern in *R. kutum* is in accordance with inflexion caudal complex, showing ontogeny of the caudal complex skeleton in agreements with its function demands during early development.

#### Key words: Tail, Ontogeny, Osteology, Deformity, Caspian kutum.

#### INTRODUCTION

Description of the anatomical development during early ontogeny of fishes is crucial for understanding the patterns behind body formation, functional trends and environmental preferences (Koumoundouros *et al.* 2001). Among the various anatomical structures, development of skeletal structures involves a series of transitions between cell proliferation and differentiation i.e. transitions from cartilage to bone (Hogan 1996; Urist 1997; Chen *et al.* 2004; Tsumaki & Yoshikawa 2005). In the skeletal structures, development of fins is important during ontogeny of fishes, since it is closely correlated with changes in swimming mode, swimming velocity, as well as feeding techniques and preferences (Kendall *et al.* 1984). From aquaculture point of view, understanding the skeletal development of commercially-important fish species can assist early diagnosis of the skeletal abnormalities, since most of the skeletal deformities are occurred during early ontogenic stages (Koumoundorous *et al.* 1997; Boglion *et al.* 2001). Skeletal deformities are crucial in aquaculture since they decrease the quality of produced fish by affecting their external morphology, growth and survivals (Hasanpour *et al.* 2016). There are many reports regarding the skeletal deformities in various fishes (Koumoundorous *et al.* 1997; Gavaya *et al.* 2002; Fraser *et al.* 2004; Sfakianakis *et al.* 2005).

Caspian kutum, *Rutilus kutum*, (Kamensky, 1901), is a commercially-important cyprinid fish in the Caspian Sea basin (Ebrahimi 2001; Salehi 2002). Since there is no information available regarding the osteological structure during ontogeny of this species, hence this study was conducted to describe the ontogeny of its caudal complex skeleton as an important locomotor function element which in turn will help to better understanding of its swimming and feeding ontogeny.

# MATERIALS AND METHODS

Larvae specimens were obtained from propagation via mass combined spawning of 15 female and 30 male broodstocks in April-May 2012, from Dr. Yousefpour Fish Hatchery Center, Siahkal, Guilan Province, Iran. The eggs were incubated in 10 L vase incubators with flow-through freshwater system at 22 °C.

After six days of incubation, eggs hatched and then transferred to a large larval collector tank (200 L). After 3 days, at the beginning of exogenous feeding, 30,000 larvae (3 larvae L<sup>-1</sup>) were transferred to a pond (0.1 ha) with a flow through freshwater system with a mean temperature, pH and DO of  $25 \pm 2.2^{\circ}$ C,  $8.1 \pm 0.5$  and  $7.4 \pm 1.1$  mg L<sup>-1</sup>, respectively. The natural water flow provided some natural prey but additional artificial feed was supplied when needed from seven-day post-hatch until the end of the experiment. The artificial feed was a specialized one designed for Caspian kutum larvae and juveniles based on a mix of protein and cereal meals.

A total of 520 larvae and fry were randomly sampled from hatching up to 50 dph (days post hatch). The sampling was performed initially from 0-20 dph every day, and then every five days up to 50 dph by sampling 20 specimens every morning from the same larval batch. Larvae were anaesthetized with 1% clove oil and fixed in phosphate buffered 5% formalin for 24 hours, then stored in 72% ethanol.

Ten specimens per each sampling time of 0-40 dph were cleared and stained according to Darias *et al.* (2010), whereas those of 45-50 dph larvae (n = 10) were cleared and stained based on the Taylor & Van Dyke (1985). The stained specimens were preserved in glycerol by adding a small piece of Thymol crystal to prevent fungi contamination (Gavaya *et al.* 2002). Osteological developments were examined using a stereoscopic dissecting microscope attached to a 13 MP Nikon camera. The cleared stained specimens were dissected and their skeletal elements were scanned by a scanner (EpsonV600) equipped with glycerol bath (Fig. 1). The skeletal elements were drawn from the obtained images using Corel DrawX5 software. Total lengths (TL) of larval specimens were measured to the nearest 0.01 mm and used as a reference point. Nomenclature and abbreviations of osteological features followed Hasanpour *et al.* (2016).



Fig. 1. Cleared and stained caudal skeleton of Rutilus kutum (50 dph, blue area, cartilage; red area, ossification).

#### RESULTS

#### **Pre-flexion stage**

At hatching, mean TL of the larvae was  $7.11 \pm 0.48$  mm lacking any caudal elements (Fig. 2A-B). The notochord was straight and primordial marginal fin-fold was present (1-4 dph). The caudal fin was begun to develop approximately at  $9.41 \pm 0.63$  mm TL (4 dph), with appearance of four cartilaginous elements beneath the notochord, including the hyporals 1-3 and parhypural, from left to right, respectively. The number of principal caudal rays (PCR) was four at 4 dph (Fig. 2C). The parhypural was developed at  $9.74 \pm 0.36$  mm TL (5 dph) as a cartilaginous rod ventral to hyporals. The numbers of PCR increased to nine at this time (Fig. 2D).

#### Early flexion stage

By the flexion stage at  $9.98 \pm 0.63$  mm TL (7 dph), the notochord showed an upward curvature posteriorly. The hypurals 4 and 5, and two haemal spines were formed. The ossification of the hypural plates 1-3 and PCR were started (Fig. 2E).

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Following formation of the epural at  $10.32 \pm 0.52$  mm TL (8 dph), the parhypural, hypurals 4 and 5 and haemal spines were ossified partially (Fig. 2F).

# Late flexion stage

The notochord flexion developed at  $11.03 \pm 0.44$  mm TL (9 dph). At this stage, the caudal centrum was comprised, the 1<sup>st</sup> preural centrum fused to the 1<sup>st</sup> ural centrum (PU1 and U1) (Fig. 2G). Along with ossification of PU1+U1 and PU 2-3, the neural spines of the preural centrums 2 and 3 were appeared at  $11.62 \pm 0.57$  mm TL (10 dph), as a part of the caudal skeleton (Fig. 2H).

# Post flexion stage

During this stage, development of the caudal complex was occurred as following:

a)  $12.89 \pm 0.17$  mm TL (13 dph): The epural cartilage was separated as neural spine of the PU1, while trace of its neural arch remained as rudimentary neural arch (RNA). The development of the cartilaginous neural spines continued (Fig. 2I).

b)  $15.44 \pm 0.67$  mm TL (25 dph): The large pleurostyle covered the anterior margin of the hypurals. A second ural centrum was appeared (Fig. 2J).

c)  $17.73 \pm 0.19$  and  $21.84 \pm 0.78$  mm TL (30 and 40 dph, respectively): The anterior half of the neural spine of the PU2 was appeared at 30 dph. This structure further developed at 40 dph. The position of the U2 was shifted anteriorly i.e. toward the PU1+U1 (Fig. 2K-L).

d)  $30.47 \pm 0.43$  mm TL (50 dph): The anterior half of the neural arch of the PU3 was appeared corresponding to development of the neural spines of the PU2 (Fig. 2M).

# DISCUSSION

The teleostean caudal complex skeleton is a well-studied structure with respect to function, homology and phylogeny (Borden *et al.* 2013). In addition, study of the anatomical development is critical in understanding the functional trends and environmental preferences of the different developmental stages (Koumoundouros *et al.* 2001). Hence, the present study provided a detailed ontogeny of the caudal complex skeleton of *R. kutum* which can be used in this regard.

Similar to *R. kutum*, one epural bone has been reported in the caudal fin of Caspian roach, *Rutilus lacustris* (Hasanpour *et al.* 2016) and *Balistes capriscus* (Matsuura & Katsurava 1985), while in other species such as *Dentax dentax* (Koumondoros *et al.* 1999) and *Cypselurus heterurus doedeleini* (Dasilao & Yamaoka 1998), three epurals present.

In addition, no fusion of the caudal skeleton elements was observed in *R. kutum*. The cyprinids have six hypural plates (Buhan 1972) but five of them had been formed till 50 dph expecting another one appear until 90 dph similar to Caspian roach (Hasanpour *et al.* 2016). Based on the results, the flexion of the notochord was occurred at 7 dph with formation of 8 cartilaginous plates beneath the notochord corresponding to the two haemal spines, parhypural and five hypurals. This pattern was similar to those of *R. lacustris* (Hasanpour *et al.* 2016) except its timing (8 dph vs. 7 dph).

Observation of the osteological development process in fish larvae and juveniles is very important and close scrutiny of each skeletal region during the rearing period is necessary to understand normal development (Park *et al.* 2016). So that, tail development contributes to enhancing improved swimming and maneuvering capability for feeding and escape from predators, whilst decreasing the energetic costs of larval movements (Osse & Van den Boogaart 2004). The allometric growth pattern of tail in *R. kutum* during larval stages is positive till 7 dph (9.98  $\pm$  0.63 mm), and isometric afterward (Ghorbanzaeh *et al.* 2014).

This external change is in accordance with our finding regarding notochord flexion. In addition, the notochord flexion corresponds with formation of the feeding apparatus, fins and gas-bladder ( $9.29 \pm 0.36 \text{ mm TL}$ ) which started along with the exogenous feeding in 5-7 dph. This reveals improvement of the movement capability in Caspian kutum after notochord inflexion (7 dph) for searching food when larvae are capable of swimming and maneuvering to catch prey and escape from predators. Notochord flexion is followed by the quick development in fin-rays and body shape, as well as a consequent improvement in locomotor ability (Hoar 1988; Koumoundouros *et al.* 1999).



**Fig. 2.** Development of the caudal skeleton of *Rutilus kutum* from 1-50 dph. (A): 1 dph, (B): 2 dph, (C): 4 dph, (D): 5 dph, (E): 7 dph, (F): 8 dph, (G): 9 dph, (H): 10 dph, (I): 13 dph, (J): 25 dph, (K): 30 dph, (L): 40 dph, and (M): 50 dph (Gray area, cartilage; orange area, ossification) (Abbreviation: EP: Epural, HS: Haemal spine, Hy: Hypurals 1-6, N: Notochord, NS: Neural spine, PH: Parhypural, Pls: Pleurostyles, PU1+U1: Compound centrum formed first preural and the first Ural centrum, PU1-2: Preural central 1,2, RNA: Rudimentary neural arch, U1-2: Ural centrum 1, 2).

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#### REFERENCES

- Boglione, C, Gagliardi, F, Scardi, M & Cataudelle, S 2001, Skeletal descriptors and quality assessment in larvae and post-larvae of wild-caught and hatchery-reared gilthead sea bream (*Sparus aurata* L. 1758). *Aquaculture*, 192: 1-22.
- Borden, WC, Grande, T & Smith, WL 2013, Comparative osteology and myology of the caudal fin in the Paracanthopterygii (Teleostei: Acanthomorpha). Global Diversity and Evolution, 419-455.
- Buhan, PJ 1972, The comparative osteology of the caudal skeleton of some North American minnows (Cyprinidae). *The American Midland Naturalist Journal*, 88: 484-490.
- Chen, W, Zhao, M & Mundy, G.R 2004, Bone morphogenetic proteins. Growth factors, 22: 233-241.
- Darias, MJ, Lan-Chow-Wing, O, Cahu, C, Zambonino-Infante, J.L & Mazurais, D 2010, Double staining protocol for developing European sea bass (*Dicentrarchus labrax*) larvae. *Journal of Applied Ichthyology*, 26: 280-285.
- Dasilao, JC & Yamaoka, K 1988, Osteological and functional development of flying fish *Cypselurus heterurus doederleini* (Teleostei: Exocoetidae). *Bulletin of Marine Sciences and Fisheries*, 18: 13-26.
- Ebrahimi, M 2001, Economic assessment of fingerling releasing in south of the Caspian Sea, Iranian Fisheries Organization, Planning and Development Office, 40 p, (in Persian).
- Fraser, MR, Anderson, TA, De-Nys, R 2004, Ontogenic development of the spine and spinal deformities in larval barramundi (*Lates calcarifer*) culture. *Aquaculture*, 242: 697-711.
- Gavaya, PG, Dinis, MT & Cancela, ML 2002, Osteological development and abnormalities of the vertebral column and caudal skeleton in larval and juvenile stages of hatchery-reared Senegal sole (*Solea senegalensis*). Aquaculture, 211: 305-323.
- Ghorbanzadeh, G, Rafiee, G, Eagderi, S, Poorbagher, H & Efatpanah, I 2014, Early development and allometric growth patterns in Caspian kutum (*Rutilus frisii kutum*) (Kamenskii, 1901). *Scientific Research Journal*, 3: 35-50.
- Hasanpour, S, Eagderi, S, Mojazi-Amiri, B & Moradi, M 2016. Skeletal development of the caudal complex in Caspian roach (*Rutilus caspicus*) (yakovlev, 1927) (Teleostei: Cyprinidae), *Biharean Biologist*, 10: 16-19.
- Hoar, WS 1988, The physiology of smolting salmonids. Fish Physiology, 11: 275-343.
- Hogan, BLM 1996, Bone morphogenetic proteins: multifunctional regulators of vertebrate development. *Genes* and Development, 10: 1580-94.
- Kendall-JR, AW, Ahlstrom, EH & Moser, HG 1984. Early life history stages of fishes and their characters, Ontogeny and Systematics of Fishes. American Society of Ichthyologists and Herpetologists, Special Publication no 1, Allen Press, Laurence, 11-22.
- Koumoundouros, G, Divanach, P & Kentouri, M 1999, Osteological development of the vertebral column and of the caudal complex in *Dentex dentex. Journal of Fish Biology*, 54: 424-436.
- Koumoundouros G, Divanach, P & Kentouri, M 2001, Osteological development of *Dentex dentex* (Osteichthyes: Sparidae): dorsal, anal, paired fins and squamation. *Marine Biology*, 138: 399-406.
- Matsuura, Y & Katsuragava, M 1985, Osteological development of fins and their supports of larval grey trigger fish, *Balistes capricus. Journal of Ichthyology*, 31: 411-421.
- Osse, J. W. M.; van den Boogaart, J. G. M., 2004: Allometric growth in fish larvae: timing and function. *American Fisheries Society Symposium*. 40, 167–194.
- Osse, J. W. M.; van den Boogaart, J. G. M., 2004: Allometric growth in fish larvae: timing and function. Am. Fish. Soc. Symp. 40, 167–194.
- Osse, JW & Van-den-Boogart, JGM 2004, Allometric growth in fish larvae: timing and function. *American Fisheries Society*, 4: 167-194
- Park, JY, Han, KH, Cho, JK, Myeong, JI & Park, JM 2016, Early osteological development of larvae and juveniles in red spotted grouper *Epinephelus akaara* (Pisces: Serranidae). *Development & Reproduction*, 20: 87-101.
- Salehi, H 2002, Economic assessment of fingerling releasing *Rutilus kutum* in Iran. *Journal of Marine Science of Iran*, 1:35-45 (in Persian).

- Sfakianakis, DG, Doxa, CK, Kouttouki, S, Koumoundouros, G, Maingot, E, Divanach, P & Kentouri, M 2005, Osteological development of the vertebral column and of the fins in *Diplodus puntazzo* (Cetti, 1777). *Aquaculture*, 250: 36-46.
- Taylor, WR & Van-Dyke, GC 1985, Revised procedures for staining and clearing small fishes and other vertebrates for bone and cartilage study. *Cybium*, 9: 107-119.
- Tsumaki, N & Yoshikawa, H 2005, The role of bone morphogenetic proteins in endochondral bone formation. *Cytokine and Growth Factor Reviews*. 16:279-85.
- Urist, MR 1997, Bone morphogenetic protein: The molecularization of skeletal system development. *Journal Bone Miner Research*, 12: 343-6.

# تکوین ساختار اسکلتی دم در ماهی سفید دریای خزر (Kamensky, 1901) (ماهیان استخوانی حقیقی: کپورماهیان) در مراحل اولیه توسعه

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# چکیدہ

مطالعه تکوین ساختار استخوانی در طی مراحل اولیه توسعه برای موفقیت در پرورش لارو ماهیان اهمیت بسیار بالایی دارد. در این مطالعه تکوین ساختار اسکلت دمی ماهی سفید دریای خزر (*Rutilus kutum*)، از روز تخم گشایی تا ۵۰ روز پس از آن بررسی شد. برای این منظور تعداد ۵۲۰ لارو و بچهماهی، به صورت تصادفی نمونهبرداری و برای مطالعه استخوانشناسی، شفافسازی و رنگآمیزی شدند. نتایج نشان داد که توسعه اسکلت دمی با ظهور هیپورالهای ۱ و ۲، در روز ۴ پس از تخم گشایی شروع شده و خمیدگی نوتوکورد در روز ۷ پس از تخم گشایی با شکل گیری هشت صفحه غضروفی در زیر نوتوکورد شامل دو خار خونی، پارهیپورال و پنج هیپورال به وقوع می پیوندد. نتایج همچنین نشان داد که ساختارهایی از قبیل کمان عصبی ابتدایی، دومین اورال و نیمهی قدامی کمان عصبی PU3 در دوره بعد از خمش از روز ۱۳ الی ۵۰ پس از تخم گشایی ظهور می یابند. نقطه عطف الگوی رشد دم در ماهی سفید دریای خزر با خمیدگی اسکلت دمی در این گونه در انطباق بوده و نتایج نشان می دهد

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